# The Role of Epithelial Markers in Breast Cancer Metastasis: Systematic Review and Meta-Analysis

## Mihaela Tzvetkova<sup>1\*</sup>, Nia Dimitrova<sup>1</sup>

<sup>1</sup>American College of Sofia, Sofia, Bulgaria \*Corresponding Author: m.tzvetkova24@acsbg.org

Advisor: Dr. Ganka Dineva, g.dineva@acsbg.org

Received July 24, 2023; Revised October 4, 2023; Accepted, October 30, 2023

### Abstract

Epithelial-mesenchymal transition is believed to be a fundamental component of cancer metastasis. Hence, epithelial markers have emerged as potential therapeutic targets and diagnostic markers of metastatic cancers, leading to their significance in cancer research. In this review, studies on 15 different markers were identified to elucidate further the role of epithelial markers in breast cancer metastasis. Based on the studies, the respective role of the epithelial markers in metastatic breast cancer was derived. The cellular mechanisms guiding the markers' behavior were investigated by identifying and describing their associated miRNAs. The studies of 5 epithelial markers that had identified cellular mechanisms affecting breast cancer metastasis were screened for undergoing meta-analysis. Twenty-one studies in total had sufficient data to undergo meta-analysis. Based on the content of the studies and the conducted meta-analysis, the results' limitations, strengths, and implications were discussed in detail. Although, due to the limited amount of studies, definite conclusions cannot be made, the meta-analysis revealed novel inferences and confirmed inferences made by other researchers on the role of the specific epithelial marker in metastatic breast cancer. Additionally, the study provides insight into significant gaps in the field and urges greater exploration of the topic.

Keywords: Epithelial markers, Breast cancer, EMT, Metastasis, miRNA

### 1. Introduction

Most instances of morbidity and mortality due to malignant tumors in women are correlated with breast cancer. Most breast cancer-related deaths occur due to metastasis, or the process by which an original primary tumor evolves to a distal secondary tumor (Hagemeister et al., 1980). Metastasis is a highly complex process that requires epithelial-mesenchymal transition (EMT) (Sun et al., 2020). During EMT epithelial cells repress their epithelial characteristics and acquire mesenchymal, as a result of changes in gene expression and gene regulation mechanisms. Since the loss of epithelial markers is associated with EMT, and hence metastatic progression, revealing their status in breast cancer metastasis can potentially reveal new therapeutic targets and biomarkers for diagnosis of metastatic breast cancer (MBC) (Tyler & Tirosh, 2021). The objective of this study is to provide a comprehensive review and meta-analysis on the role of epithelial markers in MBC, and to explore their transcriptional regulation via miRNAs. We hypothesize that specific epithelial markers play a crucial role in MBC and are influenced by distinct miRNAs, which can also serve as potential diagnostic markers and therapeutic targets.

As per the guidelines proposed by the EMT International Association (TEMTIA), we acknowledge the need of a distinct description of the cellular mechanisms guiding the role of the epithelial markers in metastasis and the contribution of genetic alterations, due to the complexity of EMT and its context dependent nature (J. Yang et al., 2020). As a result, all the epithelial markers identified in the review are associated with microRNA(miRNA), a type of transcription regulator. Not only is the inclusion of miRNAs in accordance with the guidelines proposed by



TEMTIA, but since miRNA dysregulation has been detected in multiple metastatic cancers, including breast cancer, identification of miRNA sequences and their roles can contribute to novel cancer detection techniques through their utilization as biomarkers and components of new treatments through gene editing techniques (J. Yang et al., 2020). It has been shown that certain miRNA changes can be corrected using miRNA mimics or antagomirs, normalizing the signaling pathways and the gene regulatory network, and reversing the phenotype in malignant cells (O'Bryan et al., 2017). Due to the emergence of precise gene-editing techniques such as CRISPR-Cas9, miRNAs have increased potential in cancer treatment and should be a focus of research (Godden et al., 2022). Following the TEMTIA guidelines, a necessary criterion for the markers, whose studies were subjected to meta-analysis, was clear cellular processes through which they influence MBC. Fundamental characteristics acknowledged in the analysis of the results are the molecular (luminal A/B, Triple-negative, and HER2+ enriched) and/or histological breast cancer subtypes of the samples included in the study, their tumor progression stage, and microenvironment (Q. Liu et al., 2017). The epithelial markers selected for the meta-analysis portion of the study due to their defined cellular mechanisms were B-Catenin, Nectin-4, MUC1, JAM-A, and CD44.

B-catenin is a multifunctional membrane protein that's a key component of cell-cell adhesion machinery as an intracellular signal inducer in the Wnt pathway (Shang et al., 2017). The Wnt/B-catenin signaling pathway has been shown to have a regulative role in multiple cell processes including cell motility, making its disruption a causative factor for multiple pathologies, including MBC (Komiya & Habas, 2008). In normal cells the absence of Wnt leads to the phosphorylation of cytoplasmic  $\beta$ -catenin by GSK3 $\beta$  and casein kinase I $\alpha$  (CK I $\alpha$ ), which in turn prevents nuclear accumulation of  $\beta$ -catenin, allowing its ubiquitination and subsequent degradation by the ubiquitin/proteasome system (Shang et al., 2017). Nevertheless, when Wnt binds to Frizzled (FZD), it activates Disheveled (Dsh), whose activation inhibits GSK3 $\beta$  (Zeng et al., 2008). As a result, B-catenin is not degraded and accumulates in the cytoplasm and nucleus. There, it interacts with transcription coregulators like T cell factor/lymphocyte enhancer factor (Tcf/Lef), forming a B-catenin/Lef/Tcf complex. This complex transactivates the gene that encodes cyclin D1, leading to overgrowth of cells in the lobules and ducts inside the breast (Buechel et al., 2021). Nuclear accumulation of B-catenin also results in the loss of E-cadherin and consequent loss of cell polarity and adhesion, promoting the process of EMT, and therefore metastasis (Buechel et al., 2021).

Nectins are members of the immunoglobulin superfamily (IgSF) and are components of E-cadherin-based adherens junctions in epithelial cells, thereby having a vital role in the enhancement of cellular viability and movement ability (Mandai et al., 2015). Generally, studies agree that Nectin-4 is not expressed in normal epithelium, which contributes to their increased potential to act as a biomarker or treatment in MBC. Nectin-4 has been shown to affect metastasis by modulating the CXCR4/CXCL12-LYVE-1- axis (Sethy et al., 2021). Nectin-4 overexpression leads to an increase in CXCR4 expression and LYVE-1-lymphatic vessel density (LVD). Upregulation of LVD has been associated with increased invasive abilities and poor prognosis in patients (Ramani et al., 2012). CXCR4-expressing cancer cells are attracted by CXCL12- expressing organs, thereby initiating metastasis to distant organs (Guo et al., 2016). Additionally, ADAM-17, whose expression is driven by cancer stem cells, sheds the Nectin-4 ectodomain, which interacts with endothelial Integrin-B4. This interaction promotes metastasis in breast cancer stem cells by activating the Src-PI3K-AKT-iNOS axis (Siddharth et al., 2018). In particular, Nectin-4 has been shown to promote breast cancer stem cell metastasis via the Pi3k/Akt axis through WNT/β-Catenin signaling (Siddharth et al., 2017).

JAM-A is an immunoglobulin-like molecule that acts as a tight junction protein, and as such has a role in tumor cell adhesion, polarity, invasion and migration (Severson & Parkos, 2009). The cellular mechanisms through which JAM-A affects metastasis indicate that JAM-A operates differently in tissue- and cell- specific contexts. For example, by inhibiting the Akt/B-catenin signaling pathway, JAM-A disrupts Akt-mediated phosphorylation of B-catenin, thereby preventing its accumulation in the nucleus and therefore metastasis (Nava et al., 2011). In contrast, in HER2-positive breast cancer, a type of cancer that has increased proliferation ability, increased JAM-A expression promotes HER2 expression by causing the binding of FOXA1 to the HER2 gene promoter (Cruz et al., 2022). HER2 has also been shown to activate the PI3K/Akt pathway, where PI3K phosphorylation leads to Akt2 phosphorylation, whose amplification has been associated with MBC (Milella et al., 2015). JAM-A has also been shown to activate Rap1 GTPase and β1-integrin, both of which lead to increased metastatic potential of breast tumors. Rap1 activation prohibits metastasis in other types of cancers (Yi-Lei et al., 2017).



MUC-1 is a transmembrane membrane glycoprotein associated with the protection of the epithelial layer by providing lubrication of luminal epithelial surfaces, thereby promoting motility (W. Chen et al., 2021). By interacting with ICAM-1, an adhesion receptor, glycosylated MUC-1, facilitates the interaction between epithelial and endothelial cells. This process enables adhesion of circulating cancer cells to the inner lining of the blood vessel, directly or as a result of a precedent interaction with E-selectin (Hayashi et al., 2001). Glycosylated MUC1 also interacts with Src, a non-receptor tyrosine kinases that says a key role in signal transduction pathways, thereby inducing pro-migratory Rac1- and Cdc42-dependent actin reorganization at sites of contact with endothelial cells, which promotes an invasive phenotype in the tumor cell (Shen et al., 2008). MUC1 can also drive tumor angiogenesis by upregulating vascularendothelial growth factor (VEGF), thereby promoting endothelial migration and tube formation (Khodabakhsh et al., 2021). Epidermal growth factor receptor (EGFR) stimulates growth of cancer cells, and activates STAT1 and STAT3 in breast cancer, which promote cell survival and motility. MUC1 and EGFR have a positive feedback relationship in breast cancer, resulting in dependence of EGFR prolongation on MUC1. Hence, STAT3 induces the expression of Twist one, which forms a complex with MUC-1 that results in its expression in an auto-indicative loop, accounting for its upregulation in breast cancer (Bitler et al., 2010). A subunit of MUC-1, MUC1-C, can also induce EMT and thus metastasis by activating the inflammatory NF-kB p65 pathway, which induces the transcription of ZEB1 and Bcell lymphoma 2-related protein A1 (BCL2A1) (Ahmad et al., 2009).

CD44 is a cell-surface glycoprotein involved in cell-cell interactions, adhesion, and motility, and CD44 has been used as a surface marker for breast cancer stem cells (CSCs) (Thapa & Wilson, 2016). Breast CSCs that exhibit CD44+/CD24- are potentially one of the main factors contributing to relapse of triple negative breast cancer (TNBC) due to their exacerbated self-renewal and differentiation abilities (X. Qiao et al., 2021). CD44 expression activates Rho GTPases and PI3K/AKT and MAPK-Ras, thereby promoting cytoskeletal remodeling and invasion. CD44 promotes cleavage of hyaluronan, resulting in modifications of the tumor microenvironment and essentially tumor progression. Expression of CD44 promotes docking of collagen specific MMP9. When MMP9 is found in the edges of migratory cells it promotes collagen degradation, thereby leading to an invasive phenotype, and cleavage of TGFB which also promotes invasion (Louderbough & Schroeder, 2011). Under specific conditions the ECM component hyaluronate stimulates CD44 to bind with merlin, a tumor suppressor protein, thereby conferring growth arrest in tumor cells (Herrlich et al., 2006). Other ways through which CD44 can prevent metastasis is by activating caspase-3 and hence promoting apoptosis of tumor cells, or by inhibiting PI3K activation/AKT phosphorylation (Ghatak et al., 2002).

The present meta-analysis distinguishes itself by integrating all studied epithelial markers linked to MBC and discussing their transcriptional regulation via miRNAs. We anticipated to encounter a correlation between epithelial markers and breast cancer metastasis, as well as miRNAs influence. Both miRNAs and markers could be used as therapeutic markers for targeted therapy. This approach results in a uniquely structured review, offering a more detailed depiction of the process than previously seen in other systematic reviews on the subject.

#### 2. Materials and Methods

Marker selection: All known epithelial cell markers were identified via the Bio-techne database (Epithelial Cell Markers and Intracellular Molecules, n.d.), and were assessed for role in cell adhesion through the National Library of Medicine database. To identify which of the remaining molecules are the subject of breast cancer studies, advanced search was performed via marker name, boolean operator "and", and "breast cancer metastasis". Review articles were identified through keywords: epithelial-mesenchymal transition (EMT), tumor marker, cancer metastasis, name of epithelial cell marker and were manually searched for more references on the topic. Markers implicated in more than five breast cancer metastasis studies proceeded to the next selection stage. These markers were then examined for studies exploring their connection with miRNAs. Table 1 presents the markers that play a role in MBC, their associated miRNAs, and studies discussing their involvement in MBC. The studies of markers that had identified cellular mechanisms in the context of MBC and associated miRNAs, were discussed in detail and analyzed for meta-analysis eligibility.

Meta-analysis: Odds ratio (OR) was used to examine the association between the expression of epithelial markers

and their prevalence in MBC. OR represents the likelihood of an event occurring when exposed to a specific factor, in contrast to the likelihood of the event without that exposure. In the current context, the OR offers insight into the likelihood of the occurrence of MBC in the presence of an epithelial marker, in contrast to the likelihood of MBC without the epithelial marker. A 2x2 contingency table was set up, with one axis indicating the presence or absence of MBC and the other indicating the expression or non-expression of the epithelial marker. The calculated OR was derived using the formula:  $[OR = \frac{ad}{bc}]$ , where 'a' denotes individuals with both the epithelial marker and MBC, 'b' signifies those with the marker but without MBC, 'c' represents those without the marker but with MBC, and 'd' identifies those without either the marker or MBC. In order for studies to be eligible for the meta-analysis, they should have reported their results such that marker transcription acts as a dependent variable, and variables measuring metastasis act as independent variables. +/- metastasis or +/- lymph node involvement would be seen as variables measuring metastasis. An OR value of 1 would suggest no association between the epithelial marker and MBC. In contrast, an OR greater than 1 would indicate an increased likelihood of MBC in the presence of the marker, whereas an OR less than 1 would suggest a decreased likelihood. The 95% confidence interval (CI) provides an estimation of the accuracy of the OR. A wide range of CI suggests that the OR's accuracy is low, while a narrow CI suggests greater accuracy. The 95% CI doesn't reflect statistical significance in the same manner as the p-value. However, if the 95% CI doesn't cross the null value (e.g., OR=1), it's often interpreted as evidence of statistical significance. Heterogeneity refers to the variability or differences in study outcomes. Analyzing heterogeneity offers insights into the influence of varying methodologies and conditions on the study outcomes. The heterogeneity of the data was assessed using tau square and chi square to make sure that all studies are evaluating the same effect. The tau squared (Tau<sup>2</sup>) represents between-study variance, with elevated values indicating substantial inter-study variability. The chi-square (Chi<sup>2</sup>) tests the hypothesis that the studies are evaluating the same effect, with a low p-value (typically  $\leq 0.05$ ) suggesting that heterogeneity is present beyond chance. The p-value represents the probability of observing the given data, or more extreme data, under the null hypothesis of no effect. A p-value less than 0.05 is conventionally deemed indicative of statistical significance. Furthermore, the I<sup>2</sup> statistic provides a quantification of the proportion of total variation across studies that's attributable to heterogeneity rather than chance. In the context of this research, an  $I^2$  value below 50% was interpreted as indicating satisfactory homogeneity. All statistical analyses were conducted using RevMan Version 5.

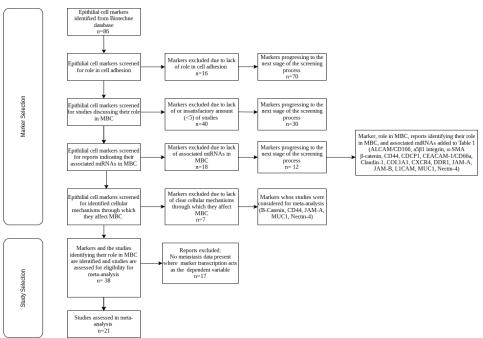


Figure 1. Flow chart representing selection procedure for epithelial cell markers and study selection for metaanalysis based on studies listed in Table 1.



### 3. Results

12 epithelial markers were shown to have a relationship with MBC in more than 5 studies, and an association with a type of miRNA, which is illustrated in Table 1. The relationship between miRNAs and metastasis was portrayed in Figure 2 and forest plots were produced to depict the statistical analysis undergone by the eligible studies of the 5 selected markers.

| Table 1 Epithelial markers associated with MBC, their role in MBC, the studies discussing their role in MBC, and |  |
|--|--|
| their associated miRNAs  |  |

| Name of Marker | Role of Marker<br>in Metastasis | Associated<br>miRNAs       | Studies discussing role of marker in metastasis   |  |
|----------------|---------------------------------|----------------------------|---|--|
| ALCAM/CD166    | tumor suppressor                | miR-125                    | (Davies et al., 2008), (Akamn et al., 2015)   |  |
| α-SMA          | oncoprotein                     | miR-200c                   | (Tang et al., 2015), (Mierke et al., 2011)  |  |
| Integrin α5β1  | oncoprotein                     | miR-31,-149                | (Wang, Yanfang, et al. 2011), (Chan, S. 2014),<br>(Augoff, K., et al 2011)  |  |
| β-catenin      | β-catenin oncoprotein           |                            | (Z. Wang et al 2015), (Nie, J. et al., 2019),<br>(Kwon, J. J. et al., 2019), (Liu, B. et al., 2018),<br>(Tan, Z. et al., 2016), (Si, W. et al., 2016) |  |
| CD44           | mixed                           | miR-205,-34a               | (Ouhtit et al., 2007) (Tse, 2005) (Zhang, Lu. et al., 2020), (Ahir, M. et al., 2020)  |  |
| CDCP1          | oncoprotein                     | miR-198                    | (Wright, H. J. et al 2017) (Hu, Y. et al., 2017)  |  |
| CEACAM-1/CD66a | tumor suppressor                | miR-342                    | (Weng, C. et al., 2016) (C. Yang et al., 2017)  |  |
| Claudin-1      | oncoprotein                     | miR-155                    | (Zhou, B. et al., 2015) (Chiang et al., 2019)   |  |
| COL1A1         | oncoprotein                     | miR-196b-5p                | (Zhu, X. et al.,2008), (Jiang, Y. et al., 2022) (W. Wu & Zheng, 2022)   |  |
| CXCR4          | oncoprotein                     | miR-9,-139                 | (Liu, Y. et al., 2021), (Cheng, CW. et al., 2021)<br>(J. Li et al., 2021)   |  |
| DDR1           | oncoprotein                     | miR-199b-p                 | (Wu, A. et al., 2018) (Baltes et al., 2020)   |  |
| JAM-A          | mixed                           | miR-495, -145              | (Cao, M. et al., 2014), (Ye, D. et al., 2019), (Naik<br>et al., 2018), (Murakami et al., 2011)  |  |
| JAM-B          | tumor suppressor                | miR-374                    | (Li, W. et al., 2019) (Bhan et al., 2013)   |  |
| L1CAM          | oncoprotein                     | miR-21-3p                  | (Doberstein, K. et al., 2014)   |  |
| MUC1           | oncoprotein                     | miR-200c, -141, -<br>1226, | (Rajabi, H. et al., 2013), (Gao, Y. et al., 2016),<br>(Kufe et al., 2010),  |  |
| Nectin-4       | mixed                           | miR-520c-3p                | (Liu, Y. et al., (2022)) (Zeindler et al., 2019),<br>(Sethy et al., 2018)   |  |



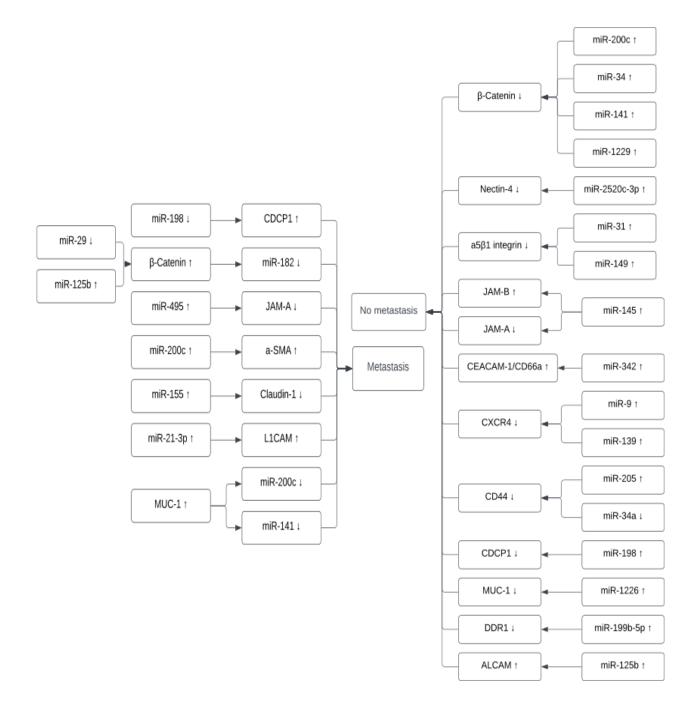


Figure 2. Description of relationship between miRNAs and epithelial markers in MBC



|  |                       |                       |                | Odds Ratio                             | Odds Ratio                                |
|--|-----------------------|-----------------------|----------------|--|---|
| Study or Subgroup  | log[Odds Ratio]       | SE                    | Weight         | IV, Random, 95% CI                     | IV, Random, 95% CI                        |
| 4.1.1 Positive   |                       |                       |                |  |   |
| Geyer 2010 14/β-catenin clone  | 0.2111 0              | .2059                 | 23.5%          | 1.24 [0.82, 1.85]                      | - <b>+</b>                                |
| Geyer 2010 17C2 clone  | 0.2237 0              | .2038                 | 23.5%          | 1.25 [0.84, 1.86]                      | +   |
| Wang LN 2015<br>Subtotal (95% CI)  | 0.2091 0              | .2732                 | 21.3%<br>68.3% | 1.23 [0.72, 2.11]<br>1.24 [0.97, 1.59] | •   |
| Heterogeneity: Tau <sup>2</sup> = 0.00; Chi <sup>2</sup> =   | 0.00, df = 2 (P = 1.0 | 0); I <sup>2</sup> =  | 0%             |  |   |
| Test for overall effect: Z = 1.68 (P   | = 0.09)               |                       |                |  |   |
| 4.1.2 High Positive  |                       |                       |                |  |   |
| Jang 2015  | 1.6275 0              | .3211                 | 19.6%          | 5.09 [2.71, 9.55]                      | <b></b>                                   |
| Lee LN 2005  | 1.2127 0              | .5781                 | 12.1%          | 3.36 [1.08, 10.44]                     |   |
| Subtotal (95% CI)  |                       |                       | 31.7%          | 4.62 [2.66, 8.00]                      | •   |
| Heterogeneity: Tau <sup>2</sup> = 0.00; Chi <sup>2</sup> =   | 0.39, df = 1 (P = 0.5 | 53); I <sup>2</sup> = | 0%             |  |   |
| Test for overall effect: Z = 5.45 (P   | < 0.00001)            |                       |                |  |   |
| Total (95% CI)   |                       |                       | 100.0%         | 1.85 [1.09, 3.13]                      | -   |
| Heterogeneity: $Tau^2 = 0.27$ ; $Chi^2 =$<br>Test for overall effect: Z = 2.27 (P<br>Test for subgroup differences: Ch | = 0.02)               |                       |                | .5%                                    | 0.1 0.2 0.5 1 2 5 10<br>Negative Positive |

Figure 3. Meta-analysis on four studies assessing the association of  $\beta$ -catenin expression with the occurrence of MBC. The pooled OR was 1.85 (95% CI: 1.09-3.13; Z=2.27; P=0.02) with heterogeneity (I2 78% P=0.0010). Two studies showed mild positive correlation between B-catenin and the occurrence of MBC: Geyer (Geyer et al., 2011) and Wang (Z. Wang et al., 2015). The subtotal OR for the two studies was 1.24 (95 % CI: 0.97-1.59; Z= 1.68(P= 0.09) without heterogeneity (I2 0% P=1). It should be noted that data from Geyer was used twice as he utilized two different types of B-catenin antibodies to prohibit metastasis. Two studies showed a high positive correlation between B-Catenin expression and occurrence of MBC: Jang (Jang et al., 2015) and Lee (Won-Lee, 2005). The subtotal OR was 4.62 (Cl% 95 2.66-8.00, Z= 2.27, P= 0.02) without heterogeneity (I2 0% P=0.53).

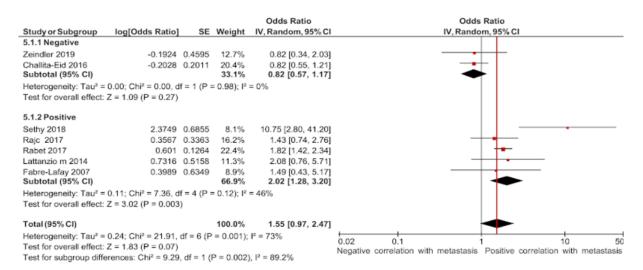


Figure 4. Meta-analysis on six studies assessing the association of Nectin-4 expression with the occurrence of MBC. The pooled OR was 1.55 (95% Cl: 0.97-2.47; Z= 1.83; P=0.07) with heterogeneity (I2 73% P= 0.002). Five studies showed positive correlation between Nectin-4 expression and the occurrence of MBC : Fabre-Lafay (Fabre-Lafay et al., 2007), Rabbit (M-Rabet et al., 2017), Lattanzio (Lattanzio et al., 2014), Sethy (Sethy et al., 2018), and Rajc (Rajc et al., 2017). The subtotal OR for the studies was 2.02 (95% Cl 1.28- 3.20 Z= 2.10; P= 0.04) with heterogeneity (I2=46%, P= 0.12). Two studies showed a negative correlation between Nectin-4 expression and the occurrence of MBC: Zeindler (Zeindler et al., 2019) and Chalita (Challita-Eid et al., 2016). The subtotal OR for those studies was 0.82 (95% Cl 0.82 Z= 1.09; P= 0.27) without heterogeneity.

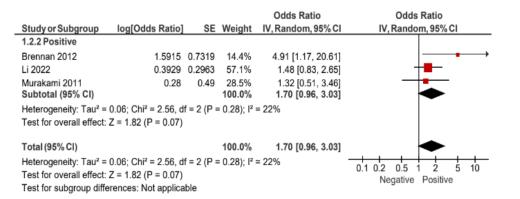


Figure 5. Meta-analysis on three studies assessing the association of JAM-A expression with the occurrence of MBC: Murakami (Murakami et al., 2011), ,Li (C.-H. Li et al., 2022), and Brennan (Brennan et al., 2013). The pooled OR was 1.70 (95% CI: 0.96-3.03; Z=1.82; P=0.28) with heterogeneity of 22% P=0.28. All three studies showed positive correlation between JAM-A and the occurrence of MBC.

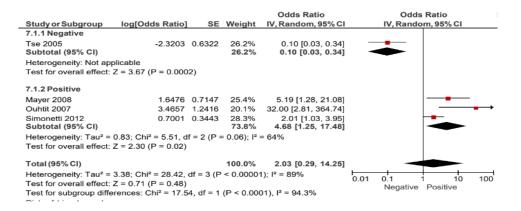


Figure 6. Meta-analysis on three studies assessing the association of CD44 expression with MBC. The pooled OR was 2.03 (95% Cl: 0.29 - 14.25; Z = 0.71; P=0.48) with heterogeneity (I2 89% P<0.00001). Three studies showed positive correlation between CD44 expression and the occurrence of MBC: Mayer (Mayer et al., 2008), Simonetti (Simonetti et al., 2012), and Ouhtit (Ouhtit et al., 2007). The subtotal OR for those studies was 4.68 (95% Cl 1.25 - 17.48; Z=2.30; P = 0.02) with heterogeneity 64%, P = 0.06. One study showed a negative correlation between CD44 and the occurrence of MBC: Tse (Tse, 2005). The subtotal OR for the study was 0.10 (95% Cl 0.10 Z= 3.67, P= 0.0002).

3.2 MUC1

| Study or Subgroup   | log[Odds Ratio]  | SE Weight       | Odds Ratio<br>IV, Random, 95% CI | Odds Ratio<br>IV, Random, 95% CI         |
|---|--|-----------------|----------------------------------|--|
| Greenberg 2003  | 3.87 1   | .15 27.6%       | 47.94 [5.03, 456.67]             |  |
| Lacunza 2010  | 0.6931 1.1   | 877 26.9%       | 2.00 [0.20, 20.51]               |  |
| McGUCKIN 1995   | 0.7909 0.3   | 188 45.5%       | 2.21 [1.18, 4.12]                | -  |
| Total (95% CI)  |  | 100.0%          | 5.03 [0.82, 30.93]               | •  |
| Heterogeneity: Tau <sup>2</sup> =<br>Test for overall effect: | 1.79; Chi <sup>2</sup> = 6.72, df = 2<br>Z = 1.74 (P = 0.08) | 2 (P = 0.03); I | 2 = 70%                          | 0.001 0.1 1 10 1000<br>Negative Positive |

Figure 7. Meta-analysis on three studies assessing the association of MUC1 expression with MBC. The pooled OR was 5.03(95% Cl: 0.82 - 30.93; Z= 1.74; P= 0.08 with heterogeneity 70% P= 0.03. Three studies showed a positive correlation between MUC1 expression and the occurrence of MBC: McGuckin (Mcguckin, 1995), Greenberg (Greenberg et al., 2003), and Lacunza (Lacunza et al., 2010).



#### **3** Discussion

Studies are unanimous in the notion that  $\beta$ -catenin expression is positively correlated with MBC and that  $\beta$ -catenin is expressed in the nucleus and/or cytoplasm of breast cancer cells. Lack of consensus occurs regarding how apparent the correlation between  $\beta$ -catenin and MBC is. As the diamond representing the subtotal OR ratio for mild positive correlation crosses the horizontal line representing the 95% interval, it's likely that Geyer and Wang do not present a statistically significant result (Fig 3). This would be in accordance with the results reported by Wang, where he stated that although high expression of B-catenin is correlated with poor patient outcome, no statistically significant correlations was noticed between B-catenin expression and metastasis (Z. Wang et al., 2015). Nevertheless, Geyer, who received similar results to Wang claimed that aberrant nuclear B-catenin expression was significantly associated with lymph node metastasis, which we failed to show in our statistical analysis, as the odds ratio for Geyer crosses the 95% interval line (Fig 3) (Geyer et al., 2011). Possible reasons include that the study only included enough data to develop an odds ratio in the context of lymph node metastasis, and not lymph vascular invasion. The lack of heterogeneity between the two studies can be attributed to both Geyer and Wang reporting their results in the context of lymph node metastasis. Factors that could have limited the results of Geyer's study include that Wang collected data in a short period of time (2006-2007), which may not be long enough to observe metastasis in patients, and that the patients were treated with anthracycline-based chemotherapy. Future longitudinal investigations could provide invaluable insights into the temporal dynamics of  $\beta$ -catenin expression and its implications for metastasis. The administration of anthracycline-based chemotherapy among patients underscores the need to rigorously examine the potential influence of such treatments on  $\beta$ -catenin expression and its subsequent association with MBC. For Lee and Jang, which indicate high positive correlation between B-catenin expression and MBC, the diamond representing the subtotal OR didn't cross the 95% interval, leading to the conclusion that the results are statistically significant (Fig 3). A potential limitation of both studies would have been the small sample sizes used. However, since the results are statistically significant, one could conclude that the small sample size doesn't significantly undermine the results. Even though the studies exhibit no heterogeneity, the subtypes of breast cancer used in the two studies are different (Jang - Sca-1 positive and Lee - ductal breast carcinoma) (Jang et al., 2015), (Won-Lee, 2005). Lack of heterogeneity despite this factor may suggest a lack of significant correlation between the role of B-catenin in MBC, and breast cancer subtype. Studying the influence of  $\beta$ -catenin expression on different breast cancer subtypes could clarify its role in metastatic potentials. Studies not included in the meta-analysis due to lack of numerical data necessary to do an odds ratio and the cellular mechanisms through which B-catenin affects cancer described in the introduction, also support the notion that B-Catenin overexpression in the nucleus and/or cytoplasm correlates with MBC (Quinn et al., 2021), (Lin et al., 2000), (De et al., 2016).

The role of Nectin-4 in MBC has been a controversial topic in research with some studies suggesting that overexpression of Nectin-4 is negatively correlated with MBC, and others proposing that it is positively correlated with MBC (Fig 4). The subtotal and individual odds ratios of the studies which suggest negative correlation between Nectin-4 expression and MBC, all cross the 95% confidence interval line, suggesting that the results are not statistically significant (Fig 4). Despite the lack of heterogeneity between the results of the two studies (Fig 4), they discuss MBC in the context of different breast cancer subtypes (Zeindler - TNBC and Chelita - ductal and lobular) and used different antibodies to locate Nectin-4 (Zeindler- AGS-22M6, ASG-22C and Chelita - M22-244b3). Since prior studies on Nectin-4 have demonstrated its sensitivity to different types of antibodies, the homogeneity between the two studies was unexpected (Lattanzio et al., 2014). A comprehensive study focusing on how various antibodies impact the detection and quantification of Nectin-4 expression would be valuable. The subtotal OR ratio of the studies representing positive correlation between Nectin-4 and MBC didn't cross the 95% confidence interval line, indicating that the results from all the studies portrayed a statistically significant positive correlation between Nectin-4 expression and MBC (Fig 4). As indicated by the crossing of the subtotal OR ratio of Rajc with the 95% confidence interval line, the study failed to report a statistically significant result (Fig. 5). Such a conclusion would be consistent with the results explicitly stated by Rajc that MBC in HER2 negative breast cancer and Nectin-4 expression are not significantly correlated with one another (Rajc et al., 2017). Even though Fabre-Lafay and Lattanzio both reported significant correlation between Nectin-4 expression and MBC in TNBC and luminal A breast cancer, respectively,



they both cross the 95% confidence interval line, indicating a statistically insignificant relationship (Fig 4)(Lattanzio et al., 2014), (Fabre-Lafay et al., 2007). Possible reasons for the difference between the results reported by them and those demonstrated by the statistical analysis include the limited amount of numerical data reported in Fabre-Lafey, and hence used in the statistical analysis, and the varied treatment the patients were subjected to in Lattanzio that could have additionally influenced metastasis. Rabet is a strong study, illustrated through its lack of cross with the 95% confidence interval line and high weight, due to its large amount of data (Fig 4). Sethy is also a strong study, with the only limitation being its heterogeneity from the other studies, likely resulting from its smaller dataset and assessment of ductal carcinomas, without specification of the molecular subtypes (Sethy et al., 2018). Except for Frabe Lafay who didn't specify the type of breast cancer carcinoma, all the other studies investigated Nectin-4 correlation in the context of a molecular subtype (Rabet - TNBC, Lattanzio and Rajc - luminal A). The inconsistency in data presentation suggests a need for standardized data collection and reporting methods to ensure comparability across studies. The study not included in the meta-analysis and the cellular processes outlined in the introduction also agree that Nectin-4 expression positively correlates with MBC (Shao et al., 2022).

The studies included in the meta-analysis all show JAM-A expression as having positive correlation with MBC. The pooled OR ratio crosses the 95% confidence interval leading to the conclusion that no statistically significant correlation between JAM-A expression and MBC can be observed (Fig 5). Murakami has a very low OR ratio meaning that the results reported aren't statistically significant, which is supported by inference made in the study (Murakami et al., 2011). Although Li reports similar results to Murakami, he states that JAM-A plays a role in several processes related to cell motility and is predominantly expressed in TNBC cells which are often associated with increased metastatic potential (Li et al., 2022). Since Li crosses the 95% confidence interval, the statistical analysis fails to reflect the reported results (Fig 5). Possible reasons include that in the study HER2 signaling and positive ER was perceived as a sign of metastasis, due to their causative relationship with TNBC. Nevertheless, to maintain homogeneity, the statistical analysis only considered the lymph node metastasis variable. A weakness of Li is the heterogeneity seen in the data pool with some patients being diagnosed in 1991. Brenan showcases a statistically significant positive correlation between JAM-A and MBC in Figure 5, supported by their own inferences in the study (Brennan et al., 2013). The heterogeneity of the data is in the acceptable range, as all the studies measure metastasis through lymph node involvement (Fig 5). Although different subtypes are used, limiting homogeneity, there are common subtypes used. For example, both Li and Brennan assess JAM-A expression in luminal A, luminal B, HER2 positive, and basal subtypes. Like Murakami, Li also assessed JAM-A expression in TNBC. The results are generally consistent with one another apart from Li who reported no correlation between JAM-A expression and MBC in basal breast cancer, whereas Brennan reported correlation between them. Brenan reported low expression of JAM-A in Luminal A breast cancer metastasis. However, Li united luminal A and B, and deduced positive correlation between JAM-A and metastasis in the subtype. More studies are needed to elucidate the role of JAM-A and metastasis in the context of luminal breast cancer subtypes. Overall, even though the three studies do not produce a statistically significant result, there are many more studies showcasing how JAM-A expression can lead to metastasis (Yang Wang and Lui, 2012), (McSherry et al., 2011) which weren't included due to lack of sufficient data to form OR ratios. There is also a study that shows how JAM-A expression can be negatively correlated with MBC (Naik et al., 2008) which we couldn't include due to the same reason. As there are explanations based on cellular mechanisms supporting both roles of JAM-A, inclusion of both types of studies would have yielded results with greater implications.

The role of CD44 has been disputed, with studies suggesting both its negative and positive correlation with MBC. The subtotal OR for the studies indicating positive correlation between MBC and CD44 doesn't cross the 95% confidence interval, meaning that the results are significant (Fig 6). However, the heterogeneity of the studies exceeds the expected range. Possible factors contributing to the heterogeneity of the results are the different ways through which MBC was measured through (Mayer - lymph node metastasis), (Ouhtit - metastasis to the liver), (Simonetti-number of invasive ductal/ micropapillary carcinomas) (Mayer et al., 2008) (Ouhtit et al., 2007) (Simonetti et al., 2012). The status of Ouhtit as an outlier can be attributed to them examining heterogeneity in vivo in mice, whereas the other two studies examined metastasis in patients. The heterogeneity among them suggests the sensitivity of CD44 to different tumor microenvironments and its context dependent nature, discussed in other studies, as well (Louderbough & Schroeder, 2011). To reduce heterogeneity, future studies could adopt a standardized method of



measuring MBC. The OR ratio of the study that reported negative correlation between CD44 and MBC, didn't cross the 95% confidence interval indicating a statistically significant result (Fig 6). A weakness of the study is the difference between the age and tumor size in the control group with no observed metastasis, and the ones with, indicating that the two variables could have affected metastasis in addition to CD44 expression. Future research should ensure that control groups are matched carefully based on factors like age, tumor size, and other relevant parameters. This would provide a more accurate assessment of CD44's role without potential confounding variables. Tse only tested for MBC in the context of standard CD44, whereas the other three studies also tested for MBC in the context of CD44 in MBC.Future research should ensure that control groups are matched carefully based on factors like age, tumor size, and other relevant parameters are such as CD44v5 and CD44v6, thereby suggesting a potential role of variants in the dual nature of CD44 in MBC.Future research should ensure that control groups are matched carefully based on factors like age, tumor size, and other relevant parameters. This would provide a more accurate assessment of CD44's role without potential confounding variables. A deeper investigation into the different CD44 variants (e.g., CD44v5 and CD44v6) and their individual or combined roles in MBC could distinguish whether certain variants have more pronounced effects on MBC than the standard CD44.More studies portraying negative correlation between CD44 would have strengthened the results, however (Lopez et al., 2005) didn't include sufficient numerical data.

Studies are unanimous in the notion that MUC1 expression positively correlates with MBC. However, as indicated by the subtotal OR ratio crossing the 95% confidence interval, the studies do not provide enough data for a statistically significant correlation (Fig 7). Although there were a total of 10 studies discussing the role of MUC1 in MBC, all leading to the conclusion stated above, we were able to find sufficient data for an OR ratio only in 3. Not only are 3 studies insufficient, but their individual sample sizes were also very small, further contributing to the statistical insignificance of the results. The data has heterogeneity in the acceptable range. However, it was unexpected that McGuckin and Lacunza have greater similarity between their results than Greenberg and Lacunza, since both Lacunza and Greenberg measured MUC1 expression in vivo, whereas McGuckin measured it in vitro (Greenberg et al., 2003) (Lacunza et al., 2010).

The data encompassed in this study facilitated the contextual interpretation of both the meta-analysis outcomes and the identification of epithelial markers implicated in MBC, highlighting the need for marker comparisons. The meta-analysis findings not only suggest new research avenues to refine our understanding of the epithelial markers' role in MBC, but also offer a critical evaluation of current literature.

#### Acknowledgment

We would like to thank Dr. Dineva for her guidance and support, which have contributed significantly to the development and completion of this review.

#### References

Ahmad, R., et al. (2009). MUC1-C oncoprotein functions as a direct activator of the nuclear factor-κB p65 transcription factor. Cancer Research, 69(17), 7013–7021. https://doi.org/10.1158/0008-5472.can-09-0523

Ahir, M., et al. (2020). Delivery of dual MIRNA through CD44-targeted mesoporous silica nanoparticles for enhanced and effective triple-negative breast cancer therapy. Biomaterials Science, 8(10), 2939–2954. https://doi.org/10.1039/d0bm00015a

Akman, H. B., et al. (2015). ALCAM is indirectly modulated by miR-125b in MCF7 cells. Tumour Biology: The Journal of the International Society for Oncodevelopmental Biology and Medicine, 36(5), 3511–3520. https://doi.org/10.1007/s13277-014-2987-5

Al-Mrabt, N., et al. (2010). Abstract P6-08-11: The potential role of junctional adhesion molecule (JAM)-2 in breast cancer cells and the expression of JAM2 in ductal mammary carcinoma. Cancer Research, 70(24\_Supplement), P6-08-11-P6-08-11. https://doi.org/10.1158/0008-5472.sabcs10-p6-08-11

Augoff, K., et al. (2011). miR-31 Is a Broad Regulator of  $\beta$ 1-Integrin Expression and Function in Cancer Cells. Molecular Cancer Research, 9(11), 1500–1508. https://doi.org/10.1158/1541-7786.mcr-11-0311



Baltes, F., et al. (2020). Targeting Discoidin Domain Receptor 1 (DDR1) signaling and its crosstalk with  $\beta$ 1-integrin emerges as a key factor for breast cancer chemosensitization upon collagen type 1 binding. International Journal of Molecular Sciences, 21(14), 4956. https://doi.org/10.3390/ijms21144956

Bhan, A., et al.. (2013). Antisense transcript long noncoding RNA (lncRNA) HOTAIR is transcriptionally induced by estradiol. Journal of Molecular Biology, 425(19), 3707–3722. https://doi.org/10.1016/j.jmb.2013.01.022

Bitler, B. G., Goverdhan, A., & Schroeder, J. A. (2010). MUC1 regulates nuclear localization and function of the epidermal growth factor receptor. Journal of Cell Science, 123(10), 1716–1723. https://doi.org/10.1242/jcs.062661

Brennan, K., et al. 2013). Junctional adhesion molecule-A is co-expressed with HER2 in breast tumors and acts as a novel regulator of HER2 protein degradation and signaling. Oncogene, 32(22), 2799–2804. https://doi.org/10.1038/onc.2012.276

Buechel, D., et al. (2021). Parsing  $\beta$ -catenin's cell adhesion and Wnt signaling functions in malignant mammary tumor progression. Proceedings of the National Academy of Sciences of the United States of America, 118(34). https://doi.org/10.1073/pnas.2020227118

Burandt, E., et al. (2014). Loss of ALCAM expression is linked to adverse phenotype and poor prognosis in breast cancer: A TMA-based immunohistochemical study on 2,197 breast cancer patients. Oncology Reports, 32(6), 2628–2634. https://doi.org/10.3892/or.2014.3523

Burkhardt, M. (2006). Cytoplasmic overexpression of ALCAM is prognostic of disease progression in breast cancer. Journal of Clinical Pathology, 59(4), 403–409. https://doi.org/10.1136/jcp.2005.028209

Cao, M., Nie, et al. (2014). MicroRNA-495 induces breast cancer cell migration by targeting JAM-A. Protein & Cell, 5(11), 862–872. https://doi.org/10.1007/s13238-014-0088-2

Challita-Eid, P. M., et al. (2016). Enfortumab vedotin antibody–drug conjugate targeting nectin-4 is a highly potent therapeutic agent in multiple preclinical cancer models. Cancer Research, 76(10), 3003–3013. https://doi.org/10.1158/0008-5472.can-15-1313

Chan, S., et al. (2014). MicroRNA-149 targets GIT1 to suppress integrin signaling and breast cancer metastasis. Oncogene, 33(36), 4496–4507. https://doi.org/10.1038/onc.2014.10

Chen, M.-J., et al. (2017). MiR-148a and miR-152 reduce tamoxifen resistance in ER+ breast cancer via downregulating ALCAM. Biochemical and Biophysical Research Communications, 483(2), 840–846. https://doi.org/10.1016/j.bbrc.2017.01.012

Chen, W., et al. (2021). MUC1: Structure, function, and clinic application in epithelial cancers. International Journal of Molecular Sciences, 22(12), 6567. https://doi.org/10.3390/ijms22126567

Cheng, C. W., et al. (2021). Mir-139 modulates cancer stem cell function of human breast cancer through targeting CXCR4. Cancers, 13(11), 2582. https://doi.org/10.3390/cancers13112582

Chiang, C. H., Hou, M. F., & Hung, W. C. (2013). Up-regulation of miR-182 by  $\beta$ -catenin in breast cancer increases tumorigenicity and invasiveness by targeting the matrix metalloproteinase inhibitor RECK. Biochimica et Biophysica Acta. General Subjects, 1830(4), 3067–3076. https://doi.org/10.1016/j.bbagen.2013.01.009

Chiang, SK., et al. (2019). DOCK1 regulates growth and motility through the RRP1B-claudin-1 pathway in claudin-low breast cancer cells. Cancers, 11(11), 1762. https://doi.org/10.3390/cancers11111762

Cruz, R. G. B., et al.. (2022). A transcriptional link between HER2, JAM-A and FOXA1 in breast cancer. Cells (Basel, Switzerland), 11(4), 735. https://doi.org/10.3390/cells11040735

# Journal of Research High School

Damonte, P., et al. (2007). EMT tumorigenesis in the mouse mammary gland. Laboratory Investigation; a Journal of Technical Methods and Pathology, 87(12), 1218–1226. https://doi.org/10.1038/labinvest.3700683

Davies, S. R., et al. (2008). Expression of the cell to cell adhesion molecule, ALCAM, in breast cancer patients and the potential link with skeletal metastasis. Oncology Reports, 19(2), 555–561. https://doi.org/10.3892/or.19.2.555

De, P., et al. (2016). Wnt-beta-catenin pathway signals metastasis-associated tumor cell phenotypes in triple negative breast cancers. Oncotarget, 7(28), 43124–43149. https://doi.org/10.18632/oncotarget.8988 Doberstein, K., et al. (2014). miR-21-3p is a positive regulator of L1CAM in several human carcinomas. Cancer Letters, 354(2), 455–466. https://doi.org/10.1016/j.canlet.2014.08.020

Doberstein, K., et al. (2014). L1CAM is expressed in triple-negative breast cancers and is inversely correlated with Androgen receptor. BMC Cancer, 14(1). https://doi.org/10.1186/1471-2407-14-958

Epithelial Cell Markers and Intracellular Molecules. (n.d.). Www.rndsystems.com. Retrieved August 1, 2023, from https://www.rndsystems.com/research-area/epithelial-cell-markers-and-intracellular-molecules

Fabre-Lafay, S., et al. (2007). Nectin-4 is a new histological and serological tumor associated marker for breast cancer. BMC Cancer, 7(1). https://doi.org/10.1186/1471-2407-7-73

Felding-Habermann, B., et al. (2001). Integrin activation controls metastasis in human breast cancer. Proceedings of the National Academy of Sciences of the United States of America, 98(4), 1853–1858. https://doi.org/10.1073/pnas.98.4.1853

Garcia-Recio, S., et al. (2022). Multiomics in primary and metastatic breast tumors from the AURORA US network finds microenvironment and epigenetic drivers of metastasis. Nature Cancer. https://doi.org/10.1038/s43018-022-00491-x

Gao, Y., et al. (2016). The roles of microrna-141 in human cancers: From diagnosis to treatment. Cellular Physiology and Biochemistry, 38(2), 427–448. https://doi.org/10.1159/000438641

Geyer, F. C., et al. (2011). β-Catenin pathway activation in breast cancer is associated with triple-negative phenotype but not with CTNNB1 mutation. Modern Pathology: An Official Journal of the United States and Canadian Academy of Pathology, Inc, 24(2), 209–231. https://doi.org/10.1038/modpathol.2010.205

Ghatak, S., Misra, S., & Toole, B. P. (2002). Hyaluronan oligosaccharides inhibit anchorage-independent growth of tumor cells by suppressing the phosphoinositide 3-kinase/akt cell survival pathway. The Journal of Biological Chemistry, 277(41), 38013–38020. https://doi.org/10.1074/jbc.m202404200

Godden, A. M., et al. (2022). An efficient miRNA knockout approach using CRISPR-Cas9 in Xenopus. Developmental Biology, 483, 66–75. https://doi.org/10.1016/j.ydbio.2021.12.015

Greenberg, R., et al. (2003). Detection of hepatocyte growth factor/scatter factor receptor (c-Met) in axillary drainage after operations for breast cancer using reverse transcriptase–polymerase chain reaction. Breast Cancer Research: BCR, 5(3). https://doi.org/10.1186/bcr588

Guo, F., et al. (2016). CXCL12/CXCR4: a symbiotic bridge linking cancer cells and their stromal neighbors in oncogenic communication networks. Oncogene, 35(7), 816–826. https://doi.org/10.1038/onc.2015.139

Hagemeister, F. B., et al. (1980). Causes of death in breast cancer a clinicopathologic study. Cancer, 46(1), 162–167. https://doi.org/10.1002/1097-0142(19800701)46:1<162::aid-cncr2820460127>3.0.co;2-b

Hattrup, C. L., & Gendler, S. J. (2006). MUC1 alters oncogenic events and transcription in human breast cancer cells. Breast Cancer Research: BCR, 8(4). https://doi.org/10.1186/bcr1515

Hayashi, T., et al.. (2001). MUC1 mucin core protein binds to the domain 1 of ICAM-1. Digestion, 63(Suppl. 1), 87–92. https://doi.org/10.1159/000051917



Herrlich, P., et al. (2006). CD44 acts both as a growth- and invasiveness-promoting molecule and as a tumorsuppressing cofactor. Annals of the New York Academy of Sciences, 910(1), 106–120. https://doi.org/10.1111/j.1749-6632.2000.tb06704.x

Hou, J., et al. (2020). The roles of integrin  $\alpha$ 5 $\beta$ 1 in human cancer. OncoTargets and Therapy, 13, 13329–13344. https://doi.org/10.2147/ott.s273803

Hu, Y., et al. (2017). miR-198 functions as a tumor suppressor in breast cancer by targeting CUB domain-containing protein 1. Oncology Letters, 13(3), 1753–1760. https://doi.org/10.3892/ol.2017.5673 Ihnen, M., et al. (2010). Expression levels of Activated Leukocyte Cell Adhesion Molecule (ALCAM/CD166) in primary breast carcinoma and distant breast cancer metastases. Disease Markers, 28(2), 71–78. https://doi.org/10.3233/DMA-2010-0685

Jang, G.-B., et al. (2015). Blockade of Wnt/β-catenin signaling suppresses breast cancer metastasis by inhibiting CSC-like phenotype. Scientific Reports, 5(1). https://doi.org/10.1038/srep12465

Jassem, J., et al. (2011). The neuronal cell adhesion molecule L1CAM as a therapeutic targe in breast cancer. Journal of Clinical Oncology: Official Journal of the American Society of Clinical Oncology, 29(15\_suppl), e21138–e21138. https://doi.org/10.1200/jco.2011.29.15\_suppl.e21138

Jiang, Y., et al. (2022). Collagen fiber features and COL1A1: are they associated with elastic parameters in breast lesions, and can COL1A1 predict axillary lymph node metastasis? BMC Cancer, 22(1). https://doi.org/10.1186/s12885-022-10092-7

Jing, X., et al. (2018). Overexpression of MUC1 predicts poor prognosis in patients with breast cancer. Oncology Reports. https://doi.org/10.3892/or.2018.6887

Kar, S., et al. (2020). Wnt/β-catenin signaling pathway regulates osteogenesis for breast cancer bone metastasis: Experiments in an in vitro nanoclay scaffold cancer testbed. ACS Biomaterials Science & Engineering, 6(5), 2600– 2611. https://doi.org/10.1021/acsbiomaterials.9b00923

Karpathiou, G., et al. (2021). Immunohistochemical analysis of L1 cell adhesion molecule and high endothelial venules in breast cancer brain metastasis. Pathology, Research and Practice, 223(153484), 153484. https://doi.org/10.1016/j.prp.2021.153484

Khodabakhsh, F., et al. (2021). Crosstalk between MUC1 and VEGF in angiogenesis and metastasis: a review highlighting roles of the MUC1 with an emphasis on metastatic and angiogenic signaling. Cancer Cell International, 21(1). https://doi.org/10.1186/s12935-021-01899-8

Kim, M. J., et al. (2020). Novel antibodies targeting MUC1-C showed anti-metastasis and growth-inhibitory effects on human breast cancer cells. International Journal of Molecular Sciences, 21(9), 3258. https://doi.org/10.3390/ijms21093258

King, J. A., et al. (2004). Activated leukocyte cell adhesion molecule in breast cancer: prognostic indicator. Breast Cancer Research: BCR, 6(5). https://doi.org/10.1186/bcr815

Komiya, Y., & Habas, R. (2008). Wnt signal transduction pathways. Organogenesis, 4(2), 68–75. https://doi.org/10.4161/org.4.2.5851

Kufe. (2010). miR-1226 targets expression of the mucin 1 oncoprotein and induces cell death. International Journal of Oncology, 37(1). https://doi.org/10.3892/ijo\_00000653

Kwon, J. J., et al. (2019). A systematic review of Mir-29 in cancer. Molecular Therapy - Oncolytics, 12, 173–194. https://doi.org/10.1016/j.omto.2018.12.011



Lacunza, E., et al. (2010). MUC1 oncogene amplification correlates with protein overexpression in invasive breast carcinoma cells. Cancer Genetics and Cytogenetics, 201(2), 102–110. https://doi.org/10.1016/j.cancergencyto.2010.05.015

Lattanzio, R., et al. (2014). Membranous Nectin-4 expression is a risk factor for distant relapse of T1-T2, N0 luminal-A early breast cancer. Oncogenesis, 3(9), e118–e118. https://doi.org/10.1038/oncsis.2014.32

Lavrsen, K., et al. (2013). Aberrantly glycosylated MUC1 is expressed on the surface of breast cancer cells and a target for antibody-dependent cell-mediated cytotoxicity. Glycoconjugate Journal, 30(3), 227–236. https://doi.org/10.1007/s10719-012-9437-7

Li, C.-H., et al. (2022). The activation of EP300 by F11R leads to EMT and acts as a prognostic factor in triplenegative breast cancers. In Research Square. https://doi.org/10.21203/rs.3.rs-1454643/v1 Li, J., et al. (2021). Vitamin D regulates CXCL12/CXCR4 and epithelial-to-mesenchymal transition in a model of breast cancer metastasis to lung. Endocrinology, 162(7). https://doi.org/10.1210/endocr/bqab049

Li, W., et al. (2022). Novel insights into the roles and therapeutic implications of MUC1 oncoprotein via regulating proteins and non-coding RNAs in cancer. Theranostics, 12(3), 999–1011. https://doi.org/10.7150/thno.63654

Li, W., et al. (2021). MicroRNA-34a: Potent tumor suppressor, cancer stem cell inhibitor, and potential anticancer therapeutic. Frontiers in Cell and Developmental Biology, 9. https://doi.org/10.3389/fcell.2021.640587

Li, Y., & Galileo, D. S. (2010). Soluble L1CAM promotes breast cancer cell adhesion and migration in vitro, but not invasion. Cancer Cell International, 10(1), 34. https://doi.org/10.1186/1475-2867-10-34

Lin, S.-Y., et al. (2000). β-Catenin, a novel prognostic marker for breast cancer: Its roles in cyclin D1 expression and cancer progression. Proceedings of the National Academy of Sciences of the United States of America, 97(8), 4262–4266. https://doi.org/10.1073/pnas.060025397

Liu, Q., et al. (2017). Factors involved in cancer metastasis: a better understanding to "seed and soil" hypothesis. Molecular Cancer, 16(1). https://doi.org/10.1186/s12943-017-0742-4

Liu, B., et al. (2018). Mir-200c/141 regulates breast cancer stem cell heterogeneity via targeting HIPK1/ $\beta$ -catenin axis. Theranostics, 8(21), 5801–5813. https://doi.org/10.7150/thno.29380

Liu, Y., Zhao, Q., Xi, T., Zheng, L., & Li, X. (2021). MicroRNA-9 as a paradoxical but critical regulator of cancer metastasis: Implications in personalized medicine. Genes & amp; Diseases, 8(6), 759–768. https://doi.org/10.1016/j.gendis.2020.10.005

Liu, Y., et al. (2022). Nectin-4 promotes osteosarcoma progression and metastasis through activating PI3K/AKT/NF-κB signaling by down-regulation of miR-520c-3p. Cancer Cell International, 22(1). https://doi.org/10.1186/s12935-022-02669-w

Lopez, J. I., et al. (2005). CD44 attenuates Metastatic invasion during breast cancer progression. Cancer Research, 65(15), 6755–6763. https://doi.org/10.1158/0008-5472.can-05-0863

Louderbough, J. M. V., & Schroeder, J. A. (2011). Understanding the dual nature of CD44 in breast cancer progression. Molecular Cancer Research: MCR, 9(12), 1573–1586. https://doi.org/10.1158/1541-7786.mcr-11-0156

Lu, L., et al. (2015). Circulating tumor cell clusters-associated gene plakoglobin and breast cancer survival. Breast Cancer Research and Treatment, 151(3), 491–500. https://doi.org/10.1007/s10549-015-3416-1

Machado-Pineda, Y., et al. (2018). CD9 controls integrin  $\alpha$ 5 $\beta$ 1-mediated cell adhesion by modulating its association with the metalloproteinase ADAM17. Frontiers in Immunology, 9. https://doi.org/10.3389/fimmu.2018.02474

# Journal of Research High School

Mandai, K., et al. (2015). Nectins and nectin-like molecules in development and disease. In Current Topics in Developmental Biology (pp. 197–231). Elsevier.

Mayer, S., et al. (2008). Increased soluble CD44 concentrations are associated with larger tumor size and lymph node metastasis in breast cancer patients. Journal of Cancer Research and Clinical Oncology, 134(11), 1229–1235. https://doi.org/10.1007/s00432-008-0397-z

McFarlane, S., et al. (2015). CD44 increases the efficiency of distant metastasis of breast cancer. Oncotarget, 6(13), 11465–11476. https://doi.org/10.18632/oncotarget.3410

Mcguckin, M. (1995). Prognostic significance of muc1 epithelial mucin expression in breast cancer\*1. Human Pathology, 26(4), 432–439. https://doi.org/10.1016/0046-8177(95)90146-9

McSherry, E. A., et al. (2011). Breast cancer cell migration is regulated through junctional adhesion molecule-Amediated activation of Rap1 GTPase. Breast Cancer Research: BCR, 13(2). https://doi.org/10.1186/bcr2853

Mierke, C. T., et al. (2011). Integrin  $\alpha$ 5 $\beta$ 1 facilitates cancer cell invasion through enhanced contractile forces. Journal of Cell Science, 124(3), 369–383. https://doi.org/10.1242/jcs.071985

Milella, M., et al. (2015). PTEN: Multiple Functions in Human Malignant Tumors. Frontiers in Oncology, 5. https://doi.org/10.3389/fonc.2015.00024

Moisini, I., et al. (2021). L1CAM expression in recurrent estrogen positive/HER2 negative breast cancer: A novel biomarker worth considering. Applied Immunohistochemistry & Molecular Morphology, 29(4), 287–292. https://doi.org/10.1097/pai.00000000000909

Morozevich, G. E., et al. (2011). Implication of integrin  $\alpha$ 5 $\beta$ 1 in human breast carcinoma apoptosis and drug resistance. Biomeditsinskaia khimiia, 57(1), 77–84. https://doi.org/10.18097/pbmc20115701077

M-Rabet, M., et al. (2017). Nectin-4: a new prognostic biomarker for efficient therapeutic targeting of primary and metastatic triple-negative breast cancer. Annals of Oncology, 28(4), 769–776. https://doi.org/10.1093/annonc/mdw678

Murakami, M., et al. (2011). Abrogation of junctional adhesion molecule-A expression induces cell apoptosis and reduces breast cancer progression. PloS One, 6(6), e21242. https://doi.org/10.1371/journal.pone.0021242

Naik, M. U., et al. (2008). Attenuation of junctional adhesion molecule-A is a contributing factor for breast cancer cell invasion. Cancer Research, 68(7), 2194–2203. https://doi.org/10.1158/0008-5472.can-07-3057

Nava, P., et al. (2011). JAM-A regulates epithelial proliferation through Akt/β-catenin signalling. EMBO Reports, 12(4), 314–320. https://doi.org/10.1038/embor.2011.16

Nie, J., et al. (2019). MiR-125b regulates the proliferation and metastasis of triple negative breast cancer cells via the Wnt/ $\beta$ -catenin pathway and EMT. Bioscience, Biotechnology, and Biochemistry, 83(6), 1062–1071. https://doi.org/10.1080/09168451.2019.1584521

O'Bryan, S., et al. (2017). The roles of oncogenic miRNAs and their therapeutic importance in breast cancer. European Journal of Cancer (Oxford, England: 1990), 72, 1–11. https://doi.org/10.1016/j.ejca.2016.11.004

Ouhtit, A., et al. (2007). In vivo evidence for the role of CD44s in promoting breast cancer metastasis to the liver. The American Journal of Pathology, 171(6), 2033–2039. https://doi.org/10.2353/ajpath.2007.070535

Peng, Y., et al. (2020). High expression of JAM2 indicates better prognosis and immunotherapy response in breast cancer. In bioRxiv. https://doi.org/10.1101/2020.12.11.421081

# Journal of Research High School

Piao, D., Jiang, T., Liu, G., Wang, B., Xu, J., & Zhu, A. (2012). Clinical implications of activated leukocyte cell adhesion molecule expression in breast cancer. Molecular Biology Reports, 39(1), 661–668. https://doi.org/10.1007/s11033-011-0783-5

Qiao, X., et al. (2021). Association of human breast cancer CD44-/CD24- cells with delayed distant metastasis. ELife, 10. https://doi.org/10.7554/elife.65418

Quinn, H. M., et al. (2021). YAP and  $\beta$ -catenin cooperate to drive oncogenesis in basal breast cancer. Cancer Research, 81(8), 2116–2127. https://doi.org/10.1158/0008-5472.can-20-2801

Rahn, J. J., et al. (2005). MUC1 mediates transendothelial migration in vitro by ligating endothelial cell ICAM-1. Clinical & Experimental Metastasis, 22(6), 475–483. https://doi.org/10.1007/s10585-005-3098-x

Rajabi, H., et al. (2013). Muc1-C oncoprotein activates the zeb1/mir-200c regulatory loop and epithelial-mesenchymal transition. Oncogene, 33(13), 1680–1689. https://doi.org/10.1038/onc.2013.114

Rajc, J., et al. (2017). Prognostic role of Nectin-4 expression in luminal B (HER2 negative) breast cancer. Pathology, Research and Practice, 213(9), 1102–1108. https://doi.org/10.1016/j.prp.2017.07.019

Ramani, P., Dungwa, J. V., & May, M. T. (2012). LYVE-1 upregulation and lymphatic invasion correlate with adverse prognostic factors and lymph node metastasis in neuroblastoma. Virchows Archiv: An International Journal of Pathology, 460(2), 183–191. https://doi.org/10.1007/s00428-011-1190-y

Sánchez-Cid, et al. (2017). MicroRNA-200, associated with metastatic breast cancer, promotes traits of mammary luminal progenitor cells. Oncotarget, 8(48), 83384–83406. https://doi.org/10.18632/oncotarget.20698

Schroeder, J. A., et al. (2003). MUC1 alters β-catenin-dependent tumor formation and promotes cellular invasion. Oncogene, 22(9), 1324–1332. https://doi.org/10.1038/sj.onc.1206291 Sethy, C., et al. (2021). Nectin-4 promotes lymphangiogenesis and lymphatic metastasis in breast cancer by regulating CXCR4-LYVE-1 axis. Vascular Pharmacology, 140(106865), 106865. https://doi.org/10.1016/j.vph.2021.106865

Sethy, C., et al. (2018). Clinical significance of Nectin-4 expression in metastasis and angiogenesis for tumor relapse. In Preprints. https://doi.org/10.20944/preprints201810.0154.v1

Severson, E. A., & Parkos, C. A. (2009). Structural determinants of Junctional Adhesion Molecule A (JAM-A) function and mechanisms of intracellular signaling. Current Opinion in Cell Biology, 21(5), 701–707. https://doi.org/10.1016/j.ceb.2009.06.005

Shang, S., Hua, F., & Hu, Z.-W. (2017). The regulation of β-catenin activity and function in cancer: therapeutic opportunities. Oncotarget, 8(20), 33972–33989. https://doi.org/10.18632/oncotarget.15687

Shao, F., et al. (2022). Nectin-4-targeted immunoSPECT/CT imaging and photothermal therapy of triple-negative breast cancer. Journal of Nanobiotechnology, 20(1). https://doi.org/10.1186/s12951-022-01444-3

Shen, Q., et al. (2008). MUC1 initiates Src-CrkL-Rac1/Cdc42–mediated actin cytoskeletal protrusive motility after ligating intercellular adhesion molecule-1. Molecular Cancer Research: MCR, 6(4), 555–567. https://doi.org/10.1158/1541-7786.mcr-07-2033

Siddharth, S., et al. (2017). Nectin-4 is a breast cancer stem cell marker that induces WNT/β-catenin signaling via Pi3k/Akt axis. The International Journal of Biochemistry & Cell Biology, 89, 85–94. https://doi.org/10.1016/j.biocel.2017.06.007

Siddharth, S., et al. (2018). The soluble nectin-4 ecto-domain promotes breast cancer induced angiogenesis via endothelial Integrin- $\beta$ 4. The International Journal of Biochemistry & Cell Biology, 102, 151–160. https://doi.org/10.1016/j.biocel.2018.07.011



Simonetti, S., et al. (2012). Immunophenotyping analysis in invasive micropapillary carcinoma of the breast: Role of CD24 and CD44 isoforms expression. Breast (Edinburgh, Scotland), 21(2), 165–170. https://doi.org/10.1016/j.breast.2011.09.004

Si, W., et al. (2016). Mir-34a inhibits breast cancer proliferation and progression by targeting WNT1 in Wnt/ $\beta$ -catenin signaling pathway. The American Journal of the Medical Sciences, 352(2), 191–199. https://doi.org/10.1016/j.amjms.2016.05.002

Sun, X., et al. (2020). Exploring the metabolic vulnerabilities of epithelial-mesenchymal transition in breast cancer. Frontiers in Cell and Developmental Biology, 8. https://doi.org/10.3389/fcell.2020.00655

Tan, Z., et al. (2016). MicroRNA-1229 overexpression promotes cell proliferation and tumorigenicity and activates Wnt/β-catenin signaling in breast cancer. Oncotarget, 7(17), 24076–24087. https://doi.org/10.18632/oncotarget.8119

Tang, X., et al. (2015). Stromal mir-200s contribute to breast cancer cell invasion through CAF activation and ECM Remodeling. Cell Death & amp; amp; Differentiation, 23(1), 132–145. https://doi.org/10.1038/cdd.2015.78

Thapa, R., & Wilson, G. D. (2016). The importance of CD44 as a stem cell biomarker and therapeutic target in cancer. Stem Cells International, 2016, 1–15. https://doi.org/10.1155/2016/2087204

Tse, G. M. K. (2005). CD44s is useful in the differentiation of benign and malignant papillary lesions of the breast. Journal of Clinical Pathology, 58(11), 1185–1188. https://doi.org/10.1136/jcp.2005.026906

Tyler, M., & Tirosh, I. (2021). Decoupling epithelial-mesenchymal transitions from stromal profiles by integrative expression analysis. Nature Communications, 12(1). https://doi.org/10.1038/s41467-021-22800-1

Vadhan, A., et al. (2022). CD44 promotes breast cancer metastasis through AKT-mediated downregulation of nuclear FOXA2. Biomedicines, 10(10), 2488. https://doi.org/10.3390/biomedicines10102488

Wang, Yanfang, et al. (2011). Integrin subunits alpha5 and alpha6 regulate cell cycle by modulating the chk1 and Rb/E2F pathways to affect breast cancer metastasis. Molecular Cancer, 10(1). https://doi.org/10.1186/1476-4598-10-84

Wang, Yang, & Lui, W.-Y. (2012). Transforming growth factor-β1 attenuates junctional adhesion molecule-A and contributes to breast cancer cell invasion. European Journal of Cancer (Oxford, England: 1990), 48(18), 3475–3487. https://doi.org/10.1016/j.ejca.2012.04.016

Wang, Yujun, Yu, L., & Wang, T. (2018). MicroRNA-374b inhibits the tumor growth and promotes apoptosis in non-small cell lung cancer tissue through the p38/ERK signaling pathway by targeting JAM-2. Journal of Thoracic Disease, 10(9), 5489–5498. https://doi.org/10.21037/jtd.2018.09.93

Wang, Z., et al. (2015). Clinical implications of  $\beta$ -catenin protein expression in breast cancer. International Journal of Clinical and Experimental Pathology, 8(11), 14989–14994.

Weng, C., Nguyen, T., & Shively, J. E. (2016). MiRNA-342 regulates CEACAM1-induced lumen formation in a three-dimensional model of mammary gland morphogenesis. The Journal of Biological Chemistry, 291(32), 16777–16786. https://doi.org/10.1074/jbc.m115.710152

Williams, K., et al. (2013). CD44 integrates signaling in normal stem cell, cancer stem cell and (pre)metastatic niches. Experimental Biology and Medicine (Maywood, N.J.), 238(3), 324–338. https://doi.org/10.1177/1535370213480714

Won-Lee, L. E. E. (2005). Prognostic Significance of Abnormal beta - catenin Expression in Breast Carcinoma. Korean Journal of Pathology, 114–119. https://pesquisa.bvsalud.org/portal/resource/pt/wpr-147993



Wright, H. J., Hou, J., Xu, B., Cortez, M., Potma, E. O., Tromberg, B. J., & Razorenova, O. V. (2017). CDCP1 drives triple-negative breast cancer metastasis through reduction of lipid-droplet abundance and stimulation of fatty acid oxidation. Proceedings of the National Academy of Sciences of the United States of America, 114(32). https://doi.org/10.1073/pnas.1703791114

Wu, A., et al. (2018). miR-199b-5p inhibits triple negative breast cancer cell proliferation, migration and invasion by targeting DDR1. Oncology Letters. https://doi.org/10.3892/ol.2018.9255

Wu, W., & Zheng, L. (2022). Comprehensive analysis identifies COL1A1, COL3A1, and POSTN as key genes associated with brain metastasis in patients with breast cancer. Evidence-Based Complementary and Alternative Medicine: ECAM, 2022, 1–7. https://doi.org/10.1155/2022/7812218

Yamashita, M., et al. (2012). Role of stromal myofibroblasts in invasive breast cancer: stromal expression of alphasmooth muscle actin correlates with worse clinical outcome. Breast Cancer (Tokyo, Japan), 19(2), 170–176. https://doi.org/10.1007/s12282-010-0234-5

Yang, C., et al. (2017). Inhibition of cell invasion and migration by CEACAM1-4S in breast cancer. Oncology Letters, 14(4), 4758–4766. https://doi.org/10.3892/ol.2017.6791

Yang, F., et al. (2019). Inhibition of Dipeptidyl peptidase-4 accelerates epithelial-mesenchymal transition and breast cancer metastasis via the CXCL12/CXCR4/mTOR axis. Cancer Research, 79(4), 735–746. https://doi.org/10.1158/0008-5472.can-18-0620

Yang, J., et al. ... On behalf of the EMT International Association (TEMTIA). (2020). Guidelines and definitions for research on epithelial–mesenchymal transition. Nature Reviews. Molecular Cell Biology, 21(6), 341–352. https://doi.org/10.1038/s41580-020-0237-9

Yao, Q., et al. (2015). CXCR4 in breast cancer: oncogenic role and therapeutic targeting. Drug Design, Development and Therapy, 4953. https://doi.org/10.2147/dddt.s84932

Ye, D., Shen, Z., & Zhou, S. (2019). function of microrna-145 and mechanisms underlying its role in malignant tumor diagnosis and treatment. Cancer Management and Research, Volume 11, 969–979. https://doi.org/10.2147/cmar.s191696

Yi-Lei, Z., et al. (2017). Roles of Rap1 signaling in tumor cell migration and invasion. Cancer Biology & Medicine, 14(1), 90–99. https://doi.org/10.20892/j.issn.2095-3941.2016.0086

Zeindler, J., et al. (2019). Nectin-4 expression is an independent prognostic biomarker and associated with better survival in triple-negative breast cancer. Frontiers in Medicine, 6. https://doi.org/10.3389/fmed.2019.00200

Zeng, X., et al. (2008). Initiation of Wnt signaling: control of Wnt coreceptor Lrp6 phosphorylation/activation via frizzled, dishevelled and axin functions. Development (Cambridge, England), 135(2), 367–375. https://doi.org/10.1242/dev.013540

Zhang, Lu, et al. (2020). miR-205/RunX2 axis negatively regulates CD44+/CD24- breast cancer stem cell activity. American Journal of Cancer Research, 10(6), 1871–1887.

Zhou, B., et al. (2015). Claudin 1 in breast cancer: New insights. Journal of Clinical Medicine, 4(12), 1960–1976. https://doi.org/10.3390/jcm4121952

Zhu, X., et al. (2018). Downregulation of MiR-196b-5p impedes cell proliferation and metastasis in breast cancer through regulating COL1A1. American Journal of Translational Research, 10(10), 3122–3132.